# Neotropical Diptera

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# Manual of Neotropical Diptera. Tabanidae<sup>1</sup>

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#### Introduction

According to Fairchild (1981: 290-291): "Adult tabanids are found in almost all conceivable habitats, from salt lakes and ocean beaches to snowline in the Andes, and from the extreme deserts of coastal Peru and Chile to the nearly perpetually rainy cloud forests of many parts of central America and the eastern slopes of the Andes. With the exception of about a dozen common and wide-ranging species which seem to prefer the more open and disturbed habiats created by man's agricultural activities, most tabanids have definite habitat preferences. This aspect of their biolgy has been little studied, although Fairchild (1953) summarized knowledge of the arboreal species and Chvala & Stary (1967) noted habitat references for Cuban speces. Philip (1978) and Fairchild (1973) noted preferences of certain species for the mangrove swamp habitat [Lutz, 1922: 148 had already noted that larvae of *Tabanus obsoletus* Wiedemann, 1821 live in that environment]. In general, Diachlorini and Pangoniini seem most restricted in choice of habitats. Chrysopsini slightly less so, while most of the ubiquitous and wide ranging species belong to Tabanini. Unlike the situation in temperate areas of the world, in the tropics adults of many species are crepuscular or even nocturnal. Sometimes this is true of both sexes, in other cases only the males appear to be active at night, judging by the results of light trap catches. This is very likely due to the generally low night temperatures prevailing in the north, but we need much more information on the periods of activity of the Neotropical species. On a seasonal basis, with the possibility of year round activity in the tropic, most species nevertheless have a definite season of flight. Lutz (1910) seems to have been the first worker in the Neotropics to note the varying flight periods of tabanids. The literature was briefly summarized by Fairchild (1942) in a report on a year's study of the seasonal distribution of 35 Panamenian species. The evidence suggested strongly that the majority of species had an annual life cycle and a definite flight season, which might be long or short and in either the wet or dry season. Very few species gave evidence of more than one brood per year, or of being on the wing throughout the year in equal abundance"

Studies on the biology, physiology and ecology of some species have been published by Allee (1926), Aragão

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(1975), Barbosa, Henriques, Rafael & Fonseca (2005), Barros & Foil (1999, 2007), Barros, Foil & Vazquez (2003), Bassi, Cunha & Coscarón (2000), Buestán (1980), Camus & Barahona (2002), Dunn (1934), Fairchild (1942f), Ferreira, Henriques & Rafael (2002), Ferreira & Rafael (2004), Foil, Leprince & Byford (1991), Galvão (1936), Gorayeb (1995, 1999, 2000), Gorayeb & Ribeiro (2001), Hack (1970), Hollander & Wright (1980), Hoppe, Dillwith, Wright & Szumlas (1990); Hine (1906), Jones (1956), Lajmanovich (1994), McKeever & French (1997), Medem (1981), Mignot & Anderson (1970), Pino, Candia, Letelier, Ostoic, Sánchez & Sánchez (1973); Poi de Neiff & Neiff (2006), Rafael (1982), Rafael & Charwood (1980), Raymond (1989), Raymond, Barre & Camus (1985), Raymond & Rousseau (1987), Roberts (1971a, 1974, 1980); Schwardt (1931), Stoffolano Jr., Angioy, Crnjar, Liscia & Pietra (1990), Stoffolano Jr. & Majer (1997), Stone (1930), Strickman & Hagan (1986), Velázquez de Ríos, Tiape Gómez, Gorayeb & Tamasauskas (2004), Wilkerson, Butier & Pechuman (1985) and Wilson (1967).

Castro (1937) published on the color of tabanid eyes and Goffe (1932) on a method of preserving and restoring the colour bands on the eyes of tabanids after death. The musculature of *Fidena (Fidena) fusca* (Thunberg, 1827) [as *besckii* Wiedemann, 1828] was studied by Barth (1961a), who also investigated the salivary glands of certain horsefly species (1961b, 1962). The mouthparts were studied by Fang, McKeever & French (1999), González, Brevis, Miranda & Sotomayor (2005), González & Flores (2004), González & Sanhueza (2003, 2004), González, Sanhueza, Flores & Vargas (2004) and MacKeever & French (1999). The possible importance in classification of the fore coxae, mesokatepimeron and mesokatespisternum was dealt with by Bernardi (1992). The wing venation was studied by Shannon & Bromley (1924). The chromosomes of certain Tabanidae has been the suject of studies by Boyes & Wilkes (1972).

Most female tabanids are hematophagous and usually require a blood meal to mature at least their eggs subsequent to the first batch. That is, they may be autogenous, partly autogenous or anautogenous. Males are exclusively nectar feeders, or do not feed, while females of a number of species seem also to be mainly or exclusively nectar feeders (Fairchild, 1981: 290) or feed upon tree sap (Roberts, 1967). Those that suck blood attack a wide range of vertebrates, mostly wild and domestic mammals and man. Some species also feed on birds (Ferreira & Rafael, 2004 (*Cairina moschata*)), alligators (Medem, 1971; Ferreira, Henriques & Rafael, 2002 (*Caiman crocodilus* (L.)), snakes (Philip, 1986 (reports *Phaeotabanus innotescens* (Walker, 1854), *Tabanus secundus* Walker, 1848 [as *T. stenocephalus* Hine, 1906], *Tabanus occidentalis* Linnaeus, 1758 [as *T. modestus* Wiedemann, 1828] and *Stenotabanus incipiens* (Walker, 1860) biting a giant anaconda in Peru); Ferreira, Henriques & Rafael, 2002 (*Eunectes murinus* (L.)) and *Tabanus vittiger* Thomson feeds on the Galapagos iguanas (Philip, 1976, 1983). A *Fidena* species from Costa Rica (Puntarenas, Rincón de Osa) has a very long proboscis, much longer than the fly's body (Merritt & Philip, 1976: 501, fig. 1).

Females are more easily captured, as they are attracted to man and animals; males are most rarely met with; for this reason, most species have been described only from females. Several kinds of traps and chemicals or the use of bait animals have been devised for the capture of horseflies, either for increasing collections or for other types of studies and control (e. g., Adkins Jr., Ezell Jr., Sheppard & Askey Jr., 1972; Ailes, Brown, Church, French & Gale, 1992; Anderson, Olkowski & Hoy, 1973; Axtell, Edwards & Dukes, 1975; Catts, 1970; Foil, 1999; Foil & Hissel, 1991; Foil & Hribar, 1995; French & Hagan, 1995; French & Kline, 1989; Granger, 1970; Hanec & Bracken, 1964; Hansens, Bosler & Robinson, 1971; Hayakawa, 1988; Hayakawa, Tanaka, Iwane & Yamashita, 1990; Hayes, Doane, Sakolky & Berrick, 1993; Hribar & Foil, 1994; Hribar, Leprince & Foil, 1991a, 1991b; Knox & Hays, 1972; Krcmar, Hribar & Kopi, 2005; Krcmar, Mikuska & Merdic, 2006; Mihok, 2002; Mizell III, Mizell IV & Mizell, 2002; Moucha, 1963; Rafael & Gorayeb, 1982; Raymond, 1987; Roberts, 1965, 1969, 1970, 1971b, 1972a, 1972b, 1975a, 1975b; Smith, Breeland & Pickard, 1965; Snoddy, 1970; Tallamy, Hansens & Denno, 1976; Thompson, 1969; Thompson & Gregg, 1971; Thompson & Holmes Jr., 1974; Thorpe & Hansens, 1978; Thorsteinson, Bracken & Hanec, 1965; Thorsteinson, Bracken & Tostowaryk, 1966; Wall & Doane Jr., 1980; Wilson, 1968; Wilson & Richardson, 1970; Wilson, Tugwell & Burns, 1966).

Adults of some species may be preyed upon by certain wasps (Cane & Miyamoto, 1979; Fischer, 1929a, 1929b; Pechuman, 1957a (*Bembix cinerea* (Handl.)) and trumpeter birds (Psophiidae) (Peres, 1996). They are also infected by *Spiroplasma* bacteria (Clark, Peterson, Whitcomb, Henegar, Hackert & Tully, 1984; French, Whitcomb, Tully, Carle, Bove, Henegar, Adams, Gasparich & Williamson, 1997). Larvae of *Tabanus nigrovittatus* Macquart, 1847 are parasitized by *Trichopria sp.* (Hymenoptera: Diapriidae) (Magnarelli & Anderson, 1980); those of *Tabanus subsimilis* Bellardi by larvae of *Macronychia sp.* near *aurata* (Coquillett) (Diptera, Sarcophagidae) (Thompson, 1978).

Some species of this family present medical and economic importance, acting as mechanical vectors of several pathogenic organisms, as viruses, bacteria, protozoans (especially *Trypanosoma*) and helminths (Barretto, 1949; Coscarón, 1998; Diéguez Fernández, Rodríguez González & Sánchez Alonso, 1997; Foil, 1989; Foil & Issel, 1991; Hack & García, 1959; Hall, Chainey, Betella & Aramayo, 1993; Hoffmann, 1961; Krinsky, 1976 (evaluates over 200 references of the disease agents transmitted by tabanids); Lutz & Núñez Tovar, 1928; Mateus, 1975; Woke, 1947). They are especially important in the mechanical transmission of trypanosomiases to some domestic and wild mammals (Bazzoli, Marques, Machado, Aquino, Alessi & Camacho, 2002; Coscarón & Led, 1968a; Curasson, 1943; Dávila & Silva, 2000; Franke, Greiner & Mehlitz, 1994; Herrera, Dávila, Novak, Abrey, Souza, d'Andrea & Jansen, 2004; Herrera, Novak, Freitas, Rademaker, Fernandes & Jansen, 2005; Horn, 1984; Lerger & Vienne, 1919; Losos, 1980; Melendez, Forlano & Figueroa, 1995; Monzón & Villavicencio, 1990; Morales, Wells & Angel, 1976; Nunes & Oshiro, 1980; Nunes, Oshiro, Dorval, Espindola, Cristaldo, Rocha & Nunes,

1994; Nunes, Oshiro, Dorval, Garcia, Silva & Bogliolo, 1993; Otte & Abuabara, 1991; Page, 1972; Paiva, Lemos, Nakazato, Mori, Bum & Bernardo, 2000; Raymond, 1990; Rodrigues, Fighera, Souza, Schild, Soares, Milano & Barros, 2005; Screck, Kline, Williams & Tidwell, 1993; Serra-Freire, 1981, 1983; Shaw, 1977; Shaw & Lainson, 1972; Silva, 2005; Silva, Barros & Herrera, 1995; Silva, Pellegrin, Lima, Ramirez & Dávila, 2004; Silva, Lima, Ramírez & Dávila, 2004; Silva, Morales, Eulert, Montenegro & Ybáñez, 1998; Silva & Silva, 2001; Silva, Silva, Schneider, Freitas, Mesquita, Mesquita, Ramírez, Dávila & Pereira, 1995, 1996; Stevens, Nunes, Lanham & Oshiro, 1989; Wells, 1984, Wells, Bethancourt & Page, 1970, ). They can also transmit bovine leukemia (Foil, Seger, French, Issel, McManus, Ohrberg & Ramsey, 1988), anaplasmosis (Hawkins, Love & Hildalgo, 1982) and the hog cholera virus (Tidwell, Dean, Combs, Anderson, Cowart & Axtell, 1972).

The larvae are elongate, of light (generally whitish) coloration, very active when excited, and given to a crawling mode of locomotion. They are found in very fast to slow running waters, in lentic habitats and in damp soils and leaf litter, usually in protected microhabitats such as under objects (rocks, logs), among algae or the roots of plants or yet among organic debris, where other invertebrates, such as oligochaetes and insect larvae, abound, upon which they feed. Some larvae are parasitized by Nematoda (Rhabditidae) worms (Camino & Stock, 1993; Stock & Camino, 1991).

The larvae leave the water to pupate and usually have a short pupal stage; they lack a cocoon and are very difficult to find.

The difficulty of finding larvae and pupae and especially of maintaining the former during the long lapse of time they require to complete their development (it may take up to three years) are probably the reason why it has been rarely possible to associate adults and preimaginal forms. Several techniques have been developed for inducing oviposition and the collection, sampling, rearing and study of larvae (e. g., Bailey, 1948; Ferreira & Rafael, 2006; Lane & Anderson, 1978; Lutz, 1920; Magnarelli, 1985; Marchand, 1917; Philip, 1928; Roberts, 1966a, 1966b, 1971, 1976a, 1976b; Shannon, 1922; Stammer, 1914; Tashiro & Schardt, 1949; Teskey, 1962; Thompson, Holmes Jr., Krauter, Raney & Clay, 1979; Thompson & Krauter, 1978; Wall & Jamnback, 1957).

Up to now, immature forms of the following neotropical species have been described:

#### Subfamily Chrysopsinae

Tribe Chrysopsini

Chrysops dampfi Philip, 1955 – Bermúdez & Bermúdez, 1994: 258, figs. 1A-E Chrysops facialis Townsend, 1897 – Burger, 1977: 192, figs. 11, 35 Chrysops flavidus Wiedemann, 1821 – Teskey, 1969: 39, fig. 31 Chrysops pachycerus Williston, 1887 – Burger, 1977: 194, figs. 12, 36 Chrysops pachycnemius Hine, 1905 – Bermúdez & Bermúdez, 1999: 259, figs. 2A-D Chrysops subcecutiens Bellardi, 1859 – Bermúdez & Bermúdez, 1999: 250, figs. 3A-E Chrysops variegatus (De Geer, 1776) – Bernúdez & Bermúdez, 1999: 261, figs. 4A-E Chrysops virgulatus Bellardi, 1859 – Burger, 1977: 195, figs. 13, 37

#### Subfamily Pangoniinae

Tribe Pangoniini

Esenbeckia (Ricardoa) delta (Hine, 1920) – Burger, 1977: 181, figs. 6, 9 Protodasyapha (Protodasyapha) hirtuosa (Philippi, 1865) – González, 1998: 466, figs. 1-14; Coscarón, 2002: 12, fig. 1F-H (larva), 15, fig. 3C (pupa)

#### Tribe Scionini

*Fidena (Laphriomyia) rufopilosa* (Ricardo, 1900) – Zillikens, Gorayeb, Steiner & Marcondes, 2005: 381 (larva, pupa, in bromeliad).

Scaptia (Scaptia) lata (Guérin-Méneville, 1835) – Coscarón & González, 1989: 251, figs. 1A-O (larva), 2A-F (pupa); Coscarón, 2002: 12, figs. 1B, D-E (larva), 15, figs. 3A-B, 5A (pupa)

#### Subfamily Tabaninae

#### Tribe Diachlorini

Acanthocera (Polistimima) vespiformis Burger, 2002 – Burger, 2002: 932, figs. 7-11

Agelanius cortesi (González & Henry, 1996 – González, 2007: 5

Agelanius fuscus González, 2004 - González, 2004a: 211, figs. 1-8

Bolbodimyia atrata (Hine, 1904) - Burger, 1977: 196, figs. 14, 38

Bolbodimyia bermudezi Tidwell & Philip, 1977 – Tidwell & Philip, 1977: 100, figs. 2a-b (pupa)

Catachlorops (Psalidia) baliopterus Gorayeb, Bermúdez, Bermúdez & Villalba, 1989 – Gorayeb, Bermúdez, Bermúdez & Villalba, 1989: 153, fígs. 4A-E (larva), 154, fígs. 5A-D (pupa)

Chlorotabans inanis (Fabricius, 1787) - Coscarón, 2002: 13 (larva), 17 (pupa)

Cryptotylus unicolor (Wiedemann, 1828) - Coscarón & Poi de Neff, 1996: 65, figs. 1-8 (pupa); Coscarón,

Mancebo & Coscarón Arias, 1998: 91, figs. 1-14 (larva), 15-20 (pupa); Coscarón, 2002: 14, figs. 1M-N, 2M-N (larva), 17, figs. 3N, 4C (pupa)

Dasybasis (Dasybasis) andicola (Philippi, 1865) – Coscarón, 1991: 10, figs. 1A-H (larva), 2A-E (pupa), 2002: 15, figs. 2C-F (larva), 19, figs. 3R, 6G (pupa)

Dasybasis (Dasybasis) bruchii (Brèthes, 1910) - González, 2002: 724, figs. 11-14, 15-21

- Dasybasis (Dasybasis) canipilis (Kröber, 1934) Coscarón, 1991: 12, figs. 3A-E (pupa), 2002: 15 (larva), 20, figs. 3S, 4L, 6H (pupa)
- Dasybasis (Dasybasis) chilensis (Macquart, 1838) Coscarón, 1991: 14, figs. 4A-D (pupa), 2002: 20, fig. 6C (pupa)
- *Dasybasis (Dasybasis) fairchildi* Coscarón & Philip, 1967 Coscarón & Philip, 1967: 45, figs. 1-15 (larva), 16-24 (pupa), Coscarón, 1991: 14, figs. 5A-B (larva), C-F (pupa), 2002: 15 (larva), 20, figs. 4G, 6E (pupa)
- *Dasybasis* (*Dasybasis*) *nigra* (Enderlein, 1925) Coscarón, 1969: 19, figs. 1 (larva), 2 (pupa), 1991: 17, figs. 6A-E (pupa), 2002: 15 (larva), 19, fig 4J (pupa)

Dasybasis (Dasybasis) nigrifrons (Philippi, 1865) - González, 2002: 273, figs. 2-4, 5-10

*Dasybasis (Dasybasis) opaca* (Brèthes, 1910) – Coscarón, 1991: 17, figs. 7A-B (larva), C-G (pupa), 2002: 15, fig. 2B (larva), 19, figs. 6A-B (pupa)

Dasybasis (Dasybasis) pruinivitta (Kröber, 1934) – González, 2004b: 1, figs. 8A-B (larva), C-J (pupa) Dasybasis (Dasybasis) testaceomaculata (Macquart, 1838) - Coscarón, 1991: 19, figs. 8A-B (larva), C-J (pupa), 2002: 15, figs. 2I-J (larva), 20, fig. 6F (pupa)

- *Lepiselaga* (*Lepiselaga*) *crassipes* (Fabricius, 1805) Lutz, 1928: 63-64 (notes); Fairchild, 1940c: 8, pl. II, figs. 1-3 (larva), 4-5 (pupa); Goodwin & Murdoch, 1974: 104, 106, figs. 22, 27; Coscarón, Coscarón Arias & Mancebo, 1996: 25, figs. 22-25 (larva), 26-32 (pupa); Coscarón, 2002: 13, fig. 11 (larva), 16, figs. 3J, 4B (pupa)
- *Leucotabanus albovarius* (Walker, 1854) Godoi & Rafael, 2007: 101, figs. 8-10, 16 (larva), 11-13, 17-18 (puparium).

Leucotabanus exaestuans (Linnaeus, 1758) - Goodwin & Murdoch, 1974

Leucotabanus flavinotum (Kröber, 1934) - Goodwin & Murdoch, 1974: 106, 108, figs. 15, 29

- Myiotabanus amazonicus Rafael & Ferreira, 2004 Rafael & Ferreira, 2004: 326, figs. 11-13 (pupa)
- Myiotabanus barrettoi Fairchild, 1971 Coscarón, Coscarón Arias & Mancebo, 1996: 21, figs. 1-10 (larva), 11-21 (pupa); Coscarón, 2002: 13, fig. 1J (larva), 16, figs. 3E, G, 4A (pupa)
- Stibasoma (Stibasoma) flavohirtum (Wiedemann, 1828) Goodwin & Murdoch, 1974: 114, fig. 33
- Stibasoma (Stibasoma) theotaenia (Wiedemann, 1828) Coscarón, Mancebo & Coscarón Arias,
- 1999: 619, figs. 6-11, 12-18; Coscarón, 2002: 14, fig. 1K (larva), 16, figs. 3L-M, 4D (pupa)

#### Tribe Tabanini

- Agkistrocerus aurantiacus (Bellardi, 1859) Burger, Martínez, Pechuman & Bermúdez, 1990: 183, figs. 2-11
- Poeciloderas quadripunctatus (Fabricius, 1805) Bermúdez & Bermúdez, 1998: 263, fig. 5; Coscarón, 2002: 17 (pupa)
- Tabanus abactor Philip, 1936 Montandon, Slosser & Lucia, 1993: 61 (larval habitat)
- *Tabanus atratus* Fabricius, 1775 Walsh, 1865 (larva), Riley, 1870 (larva), Hart, 1895 (larva), Hine, 1906: 33, fig. 10 (pupa), Stone, 1930 (larva), Jamnback & Wall, 1959 (larva), Teskey, 1969: 64, figs. 48, 114 (larva, pupa)

Tabanus boharti Philip, 1950 - Burger, 1977: 219, figs. 22, 26

Tabanus caenosus Burger, 1974 – Burger, 1977: 221, figs. 23, 47

*Tabanus claripennis* (Bigot, 1892) – Coscarón & Led, 1968b: 13, figs. 4-5 (larva), 6-7 (pupa) ; Coscarón, 2002: 14 (larva), 18, fig. 4E (pupa)

*Tabanus dorsifer* Walker, 1860–Roberts, 1962: 436, figs. 1-3 (larva), 4-6 (pupa); Burger, 1977: 226, figs. 25, 49 *Tabanus gilanus* Townsend, 1897–Burger, 1977: 228, figs. 26, 50

Tabanus laticornis Hine, 1904 – Burger, 1977: 207, fig. 18

*Tabanus lineola* Fabricius, 1794 – Hart, 1895; Philip, 1931; Schwardt, 1931: 411-412 (descr. of larva, larval period, number and duration of larval stages), 413 (descr. of pupa, the pupal period);

Tashiro, 1950; Jamnback & Hall, 1959; Teskey, 1969: 69, figs. 51, 123 (larva, pupa)

Tabanus morbosus Stone, 1938 – Burger, 1977: 236, figs. 28, 52

Tabanus nebulosus De Geer, 1776 – Coscarón, Mancebo & Coscarón Arias, 1998: 96 (as nebulosus ornativentris Kröber, 1929), figs. 21-22 (larva), 23-28 (pupa); Coscarón, 2002: 15 (larva), 18, fig. 3P (pupa) Tabanus nigrovittatus Macquart, 1847 – Teskey, 1969: 72, fig. 58 (larva, pupa)

*Tabanus oculus* Walker, 1848 – Bermúdez & Bermúdez, 1999: 264, figs. 6A-F *Tabanus platensis* Brèthes, 1910 - Coscarón, 1969: 21, fig. 4 (pupa), 2002: 18 (pupa) *Tabanus pruinosus* Bigot, 1892 – Burger, 1977: 237, figs. 29-53 *Tabanus punctifer* Osten Sackenm, 1876 – Burger, 1977: 240, figs. 30, 54 *Tabanus pungens* Wiedemann, 1828 - Coscarón, 2002: 14 (larva), 18 (pupa) (in key). *Tabanus subsimilis* Bellardi, 1859 – Thompson, 1975: 494 (larval habitats) *Tabanus triangulum* Wiedemann, 1828 – Coscarón, 1969: 21, fig. 3 (pupa), 2002: 18, fig. 4F (pupa)

Coscarón (2002) published an illustrated key to the larvae and pupae of Argentinian Tabanidae, with notes on the habitat of the larvae. González, Coscarón & Burger (1997) investigated the relationships of temperate South American Tabanidae with those from the Australasian region.

#### Key to Subfamilies

- 1. Tergite 9 undivided in both sexes (Figs. 1F, tix, 2H, K, 3H, L, 4I, L, 7H, 8C, 9I, 10I, 11I, 12H, 13I, 15G, 16L, 17H, 20H, L, 22F, 23E, 24C, J, 25G, K, 26D, 30I, 32G, 33F, 34E, 35D). Antennal flagellum generally with 8 (or 7) distinct flagellomeres (Figs. 2A-B, 3A-B, 4N, 5A-B, 6A, C, 7D, 8A, 9G, 10C, 20C, 22D, 23C, 25D, 28C, 35A, 37A, C, 38D) or with a basal plate of fused flagellomeres (Figs. 11D, 12C, 13C, H, 15F, J, 16C, 17C, 18C, 21B). Gonostylus of male terminalia simple, pointed (Figs. 1D, 2L-M, 3G, 4H, 24D-E, N, 25N, O, 31B-C, 33C-D, 36C-D) or double (bifid) (Figs. 8D, E, 10F-H, 12E-F, 13J, 15H-I, M, 17E-G, 20J-K, 23F-G). Female with caudal end of spermathecal ducts simple, without cup-like expansion (Figs. 1E, 2G, 3K, 4K, 7F, 9H, 11G, 13H, 16J, 18F. 22G, 24I, 25L, 26F, 27C, 29F, 30J, 32F, 35F). Ocelli and hind Tergite 9 divided, forming two separate plates in both sexes (Figs. 39F, 40E, 43G, 46F, 48H, 49E, 50J, 51F, N, 52I, 55M, 56I, 57F, 60J, 64G, 65E, 66G, 67G, 70B, 71J, 74G, 77I, 78H, 79J, S, 81F, 82C, 83F, 84H, 85L, 86D, 87H, 88D, 90G, 91D, 92G, 93I, 94G, 95F, 96G, 97I, 98H, 101J, 102G, 103I, 104E, 105D, 106G, 109H). Antennal flagellum with a basal plate plus 4 or fewer flagellomeres (Figs. 39B, 41G, M, 43B, 44A-B, 45D, 47C, 48C, 50D, 51B, I, 52C, O, 53A-B, 54A-B, D-E, G, 55B-C, J, 56C, 58B, F, I, L, 59D, H, 60D, 61B, 62H, 63C, 64C, 66C, 67C, 68A-B, 69C, 73A-B, 74B, 75A, 76A, 77A, C, 78C, K, 79D, M, 80C, 81B, 83B, 84C, 85C, 86A, 87B, 88A, 89E, 91B, 92C, 93D, 94B, 95A, C, 96B, 97C, 98B, 99B, 100B, 101D, 102C, 103C, 106B, 107D, 108C, 109B-C). Gonostylus of male terminalia either as above or truncate at apex. Female with caudal ends of spermathecal ducts simple or with cup-like expansion. Ocelli and hind tibial spur present or absent. Eye

#### Subfamily Pangoniinae (Figs. 1-38)

#### Key to tribes

- Male terminalia with dististylus bifid (Figs. 8D-E, 10F-H, 12E-F, 13J, 15H-I, M, 17E-G, 20J-K, 23F-G). Eyes generally bare (pilose in *Veprius* (Figs. 11A-B), *Austromyans* (Fig. 6A), *Protodasyapha* (Figs. 10A-B) and *Fairchildimyia* (Fig. 14A)). Face not markedly conically produced, the proboscis rarely much longer than head height (Figs. 5A-B, 6A, 7A-B, 9A-B, 10A-B, 11A-B, 12A-B, 13B, F-G, 14A, 15A-B, 16A-B, 17A-B, 18A, 19A-C, 20A-B, 21A, 22A-B, 23A-B). Vein R<sub>4</sub> nearly always with a strong appendix (stump vein, reactivation of R<sub>3</sub> field) (Figs. 3C, 5E, 15C, 18E, 19D) .... 3

#### Tribe Mycteromyiini (Figs. 1-4)

#### Key to genera

1. Male terminalia with gonocoxite bearing tuberosities in the (Fig. 1D). Female: tergite 10 undivided (Figs. 1F <i>t x</i> , 2F (Fig. 1F, 2G) (Chile)	median internal surface (Fig. 1D <i>tu</i> ). Aedeagus funnel-shaped I); caudal spermathecal duct with sclerotized portion "twisted" <i>Promycteromyja</i> Coscarón & Philip 1979
Male terminalia with gonocovite without tuberosities in the	median internal surface (Figs 3G 4H) Aedeagus somewhat
iviale terminana with gonocoxite without tuberosities in the	incutati internat surface (11gs. 30, 411). Acucagus soinewhat
thicker distally, not funnel-shaped (Figs. 3G, 4H). Fe	male: tergite 10 divided (Figs. 3L, 4L); caudal spermathecal
ducts sclerotized portion not "twisted" (Figs. 3K, 4K)	
2(1). Large flies (wing length 14-17mm). Male gonocoxite with distally (Fig. 3H). Female: caudal spermathecal ducts 3K) (Chile)	th thin and elongated apical style (Figs. 3G, 4H) and cerci acute relatively thin, with more or less thumb-like diverticles (Fig. <i>Mycteromyia</i> Philippi, 1865
Madium to small fligs (wing length & 11mm) Male: games	tulue with thick and chart anical projection (Fig. 44); corei
Medium to sman mes (wing lengur 6-1 mini). Male. gonos	tytus with thick and short apical projection (Fig. 4H), cerci
truncated distally (Fig. 4I). Female: caudal spermat	hecal ducts thick, with constrictions (Fig. 4K). (Argentina)

Silvestriellus Brèthes, 1910

#### Tribe Pangoniini (Figs. 5-21)

#### Key to genera and subgenera

Palpus without sensorial groove; if a sensory organ present, then its opening is a circular or elliptical orifice. Eye bare or pilose. Antennal flagellum with variable number of free flagellomeres, with or without a basal plate. Frons and proboscis variable
2(1) Frons with a well evident, bright callus (Figs. 5B-C)
Frons without callus, or, if a callus present, not very evident and with rugosities
3(2). Frons vey wide, ratio length/width 1.1. Body in both sexes unusually pilose, hiding the palpi and the mouthparts (U.S.A. (California), Mexico (Baja California Norte))
Frons moderately wide, ratio length/width 1.5-1.7. Body never with abundant pilosity, which does not hide the palpi and the mouthparts
<ul> <li>4(3). Callus sub-quadrate, touching the eyes, ratio length/width 1.5 (Figs. 5A, C). Gonostylus with a basal process (Canada, U.S.A. (Arizona, California, Montana), Mexico)</li></ul>
<ul> <li>5(2). Eyes hairy. Body very pilose. Frons with callus; callus elongated, not touching the eyes and nearly reaching ocellar tubercle (Mexico (Baja California))</li> <li>Brennania Philip, 1935</li> <li>Eyes bare (except in some males). Frons without callus</li> </ul>
6(5). Male eyes with hairs. Gonostylus with basal process pointed outwards (Canada to Mexico) <i>Stonemyia</i> Brennan, 1935 Male eyes bare. Gonostylus without an outwardly projected basal process (U.S.A. (California), Mexico (Baja California)) 
<ul> <li>7(1). Eye distinctly pilose (Figs. 6A, 10A,11A-B, 14A, 15A-B, 16A-B, 17A-B). Proboscis short, not much longer than palpus, the labella large and fleshy (Figs. 6A, 7A-B, 9A-B, 10A-B). Frons broad (Figs. 7B-C, 9A, C), seldom over twice as high as wide, usually with median or basal calli</li></ul>
8(7). Antennal flagellum with 8 flagellomeres (Figs. 6C, 7D, 8A, 9G, 10C). Palpus apically without orifice of sensory organ 9 Antennal flagellum with a consolidated basal plate composed of fused flagellomeres (Figs. 11D, 12C, 13C, 15F, J, 16C, 17C). Palpus apically with orifice of sensory organ (generally)
9(8). Large flies (body length 15mm). Frons without callus (Figs. 6A-B). Sternite 8 with narrow base (Fig. 6E) (Central Chile) 
Medium-sized flies (body length 9-12mm). Frons with callus (Figs. 7A-C). Sternite 8 wider basally (Figs. 7G, 9J) (genus <i>Protodasyapha</i> Enderlein, 1922)
10(9). Frons about as high as wide at base (Figs. 7B-C). Frontal callus wide, high in the middle, touching the eyes (Figs. 7B-C). Female: clypeus without dark spot; basal portion of spermathecal ducts strongly sclerotized and apical portion weakly sclerotized (Fig. 7F); gonapophyses with a weak median notch (Fig. 7G). Male gonostylus with the dorsal branch weakly curved and of same length as the ventral branch (Figs. 8D-E) (Central Chile)
Frons higher than wide, frontal callus elongate, flat, not touching eyes (Figs. 9A-C). Female: clypeus with dark spot (Figs. 9A-B), basal portion of spermathecal ducts weakly sclerotized and apical portion strongly sclerotized (Fig. 9H); gonapophyses with a deep median notch (Fig. 9J). Male gonostylus with dorsal branch hardly curved and longer than ventral branch (Figs. 10F-H) (Central Chile and central-west Argentina) Protodasyapha (Curumyia) Coscarón, 1976
<ul><li>11(8). Blackish flies, wing not fumose, no dark spot on crossveins. Apical segment of palpus without sensorial orifice 12</li><li>Greyish to greyish-brown flies, wings fumose on crossveins and sometimes with a peculiar pattern (Fig. 15C). Apical segment of palpus with sensorial orifice</li></ul>

12(11). Frons without callus, only two elongated dark spots present (Figs. 11A, E). Basal plate of flagellum with evident transverse sulci (Figs. 11D, 12C). Female: distal spermathecal ducts bulbous (Fig. 11G). Male gonocoxite wider at

base and gonostylus with dorsal branch narrower apically than ventrally (Figs. 12E-F) (Central Chile and southwestern
Argentina)
Frons with evident calli (Fig. 13A). Basal plates of flagellum without transverse sulci (Figs. 13C, H). Female: distal spermathecal
ducts with similar diameter from base to apex (Fig. 13E). Male gonocoxite narrow at base and gonostylus branches
of approximately same size (Fig. 13J) (Mexico (Baja California))

16(15). Proboscis short, markedly shorter than head height (Figs. 19B-C)	. 17
Proboscis longer than head height (Figs. 19A, F, 20B)	. 18

18(16). Proboscis entirely sclerotized, narrow, forceps-like, apparently without pseudotracheae on inner surface (Fig. 19A). Palpus long, slender to broad. Wing at most slightly tinged. Hairs on oculogenal margin sparse (Panama to Argentina) .... *Esenbeckia (Proboscoides)* Philip, 1943

Labella broader, partly or wholly sclerotized and with pseudotracheae on inner aspect. At least a single row of long hairs along oculogenal margin, immediately below sub-antennal suture. Palpus slender lo spathulate or pointed (Figs. 20D-E), at most flattened on outer aspect, generally over half length of proboscis (Mexico to Argentina) ...... *Esenbeckia (Esenbeckia)* Rondani. 1863

#### Tribe Scepsini (Figs. 22-23)

The tribe includes the sole genus *Scepsis* Walker, 1850 (Figs. 22A-H, Q; 23A-G,  $\sigma$ ), distributed along seashores of southern Brazil and Uruguay.

#### Tribe Scionini (Figs. 24-38)

#### Key to genera and subgenera

3(2). Antennal flagellum with 8 clearly separated flagellomeres (Figs. 25D, J). Palpus flattened or grooved outwardly (Figs. 25C, E-F) (Andean area, from Peru and Bolivia to southern Argentina and Chile) ... Scaptia (Scaptia) Walker, 1850 Antennal flagellum with basal flagellomeres partially fused, forming a pseudoplate and 4 to 6 free flagellomeres (Fig. 28C). Palpus stout and cylindrical (Fig. 28D). Small, densely haired flies (Southern Chile) ..... Scaptia (Pseudomelpia) Enderlein, 1922

8(7). Cell r<sub>5</sub> broadly open (Fig. 32E). Face pollinose and with abundant long hairs (Fig. 32A). Small flies, usually with strongly patterned mesonotum (Fig. 32D) and long and slender proboscis (Fig. 32A) (Southeastern Brazil) ....... *Fidena (Neopangonia)* Lutz, 1909

Cell r <sub>5</sub> normally closed, always coarctate. Face pollinose or bare, rarely with sparse, scattered, long hairs (Figs. 33A, 34A, G, 35A, 36A). Mesonotum rarely strongly patterned
9(8). Hind tibia and all femora with long, dense, outstanding hairs (Fig. 33D). Face largely or wholly bare (Fig. 33A) (Peru, Brazil, Bolivia)
All tibiae sparsely short-haired. Femora short or long-haired. Face bare or pollinose
<ul> <li>10(9). Palpus inflated, shining, grooved on outer aspect (Fig. 34C). Frons with a small subbasal protuberance (Figs. 34A-B). Face pollinose. Cell r<sub>5</sub> long-petiolate (Northwestern Argentina)</li></ul>
<ul> <li>11(6). Flagellomeres without projections, the first flagellomere much enlarged and densely haired dorsally (Figs. 37B) (Amazon basin of Ecuador, Brazil and Bolivia)</li></ul>
12(11). First flagellomere with long, hirsute, forwardly projected, dorsal, finger-like horn (Figs. 38A, D); second to sixth flagellomeres with shorter teeth, seventh and eighth flagellomeres fused into a single, long, cylindrical segment (Figs. 38A, D) (Northern South America)
First to sixth flagellomeres with long dorsal and ventral finger-like processes, those on the first longest, and then decreasing progressively in size towards apex; seventh flagellomere very short, often fused to long, finger-like eighth flagellomere (Fig. 37A) (Panama to Ecuador)

## Subfamily Chrysopsinae (Figs. 39-44)

#### Key to tribes, genera and subgenera

<ol> <li>Antennal scape nearly as long as wide and pedicel half as long as wide (Figs. 39B). Antenna shorter than anteroposterior thickness of head (Fig. 39C). Frons elongated, about 3 times as long as wide at base (Fig. 39A). Eye with a narrow transverse band (Fig. 39C). Tabaninae-like flies. Female: genital furca not elongated basally (Fig. 39D). Male gonostylus thick apically (Fig. 39G). (Chile) (Tribe Bouvieromyiini)</li></ol>
2(1). Eye hairy, without spots (Fig. 40A). Body slender, abdomen basally constricted, wasp-like. Wing blackish, more or less homogeneously pigmented; cell r <sub>5</sub> closed (Fig. 47B). (Amazonian region of Brazil) (Tribe Rhinomyzini) Betrequia Oldroyd, 1970
Eye bare, with peculiar spots (Figs. 41A, E, K). Body never as above. Wing from hyaline to (generally) with specific spots or bands and cell r <sub>5</sub> open (Figs. 41C, H, N, 43C). Female: genital furca basally with outwardly directed small projections (Figs. 41J, P, 43E) (Tribe Chrysopsini)
3(2). Eye in life irregularly-speckled. Wing hyaline or with clouds on crossveins and elsewhere, but not with distinct crossbands (genus <i>Silvius</i> Meigen, 1820)
Eye in life specifically and characteristically patterned with spots and bands (Figs. 41A, E, K). Wing nearly always with dark cross band (if cross band absent, then abdomen globose and spotted, or slender, black and shining) (Figs. 41C, H, N). Male terminalia as in Fig. 42. (Cosmopolitan)
<ul> <li>4(3). Antennal flagellum longer than scape and pedicel together (Fig. 43B). Flies with predominantly grey abdomen (Fig. 43D) and strongly spotted wing (Fig. 43C). R<sub>4</sub> usually without stump vein at fork (Fig. 43C). (Southern and western USA to Guatemala)</li></ul>
Antenna much longer than head width the flagellum clearly shorter than pedicel and not over half length of scape (Figs.

#### Subfamily Tabaninae (Figs. 45-109)

#### Key to tribes

#### Tribe Diachlorini (Figs. 45-104)

#### Key to genera and subgenera

- Without either of the above characters; basal plate at most obtusely angled dorsally and the labella wholly pollinose .... 29

- 5(4). Vein R<sub>4</sub> bent abruptly forward, so that cell r<sub>2+3</sub> is somewhat narrowed at wing margin (Fig. 45F). All tibiae more or less inflated. Callus nearly as wide as frons and prolonged to the upper border (Figs. 45A, C). Frontoclypeus not entirely bright. Male terminalia as in Figs. 46A-B. (Neotropical, except southern area) ...... Bolbodimyia Bigot, 1892

- 8(7). Very small (wing length 6.5-7.0mm), muscoid-like flies, with striped thorax and banded abdomen (Fig. 51K). Frontal callus clavate; subcallus bare on middle (Fig. 51A). Eye with a single dark median stripe (Figs. 51A, G-H). Palpus very short and inflated, hardly half length of slender proboscis (Figs. 51C, J), latter with small, compact, partly

9(2). Metallic blue flies with densely pilose eye, transverse swolle	en frontal callus (Fig. 52N), inflated bare subcallus and
gena, and antennal flagellum with dorsal projection nearly	reaching apex of basal plate (Fig. 52O). Wing hyaline
(Ecuador)	Eristalotabanus Kröber, 1913
Without such a combination of characters. Never metallic blue	

Tibiae inflated, or tubercle at vertex absent, or palpus inflated, or dorsal flagellar tooth clubbed, or very short, or frons broad .... 25

20(19). Cell r<sub>5</sub> closed or strongly coarctate (Figs. 59F, J). Wing with dark pattern leaving area around apices of basal cells or most of discal cell and apex, or oval spots on r<sub>1</sub> and r<sub>3</sub>, clear or paler (Figs. 59F, J). Frons very narrow, the callus ridge-like (Figs. 59A-B, G). Antennal flagellum with long slender projection, its apex often curved (Figs. 59D, H). Labella wholly sclerotized. Palpus very slender (Figs. 59A, E, I). (Mexico to Brazil) ..... *Catachlorops (Psalidia)* Enderlein, 1922 Cell r<sub>5</sub> open; if somewhat coarctate, wing never patterned as above (Figs. 60F, 61C, 62B). Other combinations of characters .... 21

	1	(	1 /	,
Not as above				22
			•••••	

22(21). Wing with definite pattern of bands or spots (Figs. 61C, 62B)	
Wing without a definite pattern; yellow, smoky, or veins margined with brown	24

apex of discal cell, tip of  $R_{2+3}$  and  $R_4$ , and bases of cells  $r_1$  and  $r_5$  or only two connected dark patches (Fig. 62B). Frons generally wider, callus clavate, rarely obsolescent (Figs. 62A, C-E). Labella largely pollinose. Thorax not striped. Abdomen without prominent median white triangles on tergites 1-6 (North America and tropical South America) ... *Catachlorops (Psarochlorops)* Fairchild, 1969

- Frontal callus reduced to a short, narrow ridge, small streak, or virtually absent (Fig. 66B). Wing glass clear to fumose, the coastal cell yellowish. Yellow, greenish, or brown unicolorous flies, the legs unicolorous, the body without contrasting hair patterns. Antenna with robust basal plate (Figs. 66C, 67C). Female: genital furca with acuminate branches basally (Fig. 66E); sternite 8 with base approximately as wide as gonapophyses (Fig. 66H); cerci subcircular (Fig. 66G). Male terminalia as in Figs. 67E-G. (Panama to Argentina, except southern area) ........... Cryptotylus Lutz, 1909

- times as long as wide (Fig. 71E). Body without greenish scales (South America to central Argentina)

..... Lepiselaga (Conoposelaga) Barretto, 1949

 32(31). Wing with strong, solid, black areas including at least basal half of wing and contrasting with remainder of wing. Female genital furca without projections basally (e. g., Fig. 74E)
 33 Not as above

- Frons less narrow, frontal index = 5.8. Frontal callus subtriangularly shaped, not touching the eyes. Basal flagellomere with relatively elongated process (Fig. 75A). Distal angle of wing not darkened and with 1 + 1 light spots on median distal darker area (Fig. 75B). Female sternite 8 and gonaphyses hardly wider than long (Fig. 75C). Genital furca narrower basally (Fig. 75E). Male terminalia as in Figs. 75F-G (Brazil: Rio de Janeiro to Santa Catarina) ...... *Stigmatophthalmus* Lutz, 1913
- 35(32). Notopleural lobes bluntly conical, protuberant (Fig.76A). Fore tibia inflated, hind tibia flattened and with a fringe of long hairs. Subcallus bare, frontal callus broadly clavate (Fig. 76B). Wing with a narrow abbreviated dorsal band below stigma (Fig. 76C). Eye unicolorous. Female: genital furca basally with two acuminate divergent branches (Fig. 76D), sternite 8 at base narrower than gonapophyses (Fig. 76E) (Ecuador, Peru, Brazil: Amazonas) ... *Eutabanus* Kröber, 1930 Notopleural lobes not as above, tibiae usually slender, the hind pair never flattened. Female: genital furca generally not projected into elongated branches (Figs. 77F, 78F, 79H, 83D, 84F, 85G, 87E, 92F). Sternite 8 at base generally as wide

39(38). Subcallus and face largely bare and shining. Wing glass clear, the stigma yellow. Eye with slender green lines forming two narrow transverse loops. Scutellum contrastingly pale haired, the abdomen black, shining, immaculate. All tibiae largely white
At least face wholly pollinose. Eye not as above, with two or more wider green bands (Figs. 79B-C). Other combinations of characters
40(39). Wing glass clear, the stigma yellow. Eye with slender green lines forming two narrow transverse loops (Brazil)
Wing hyaline, with a subapical costal brown patch. Eye with two iridescent dark blue stripes on a black background. (Colombia)
41(39). Frons distinctly widened below, the callus large, protuberant, filling width of frons. Wing smoky, veins heavily margined, appendix at fork of $R_4$ long. (Brazil (Rio de Janeiro to Santa Catarina))
<i>Stenotabanus (Melanotabanus)</i> Lutz & Neiva, 1914 Frons parallel-sided or narrowed below. Other combinations of characters
42(41). Frons narrow inferiorly, over twice as wide at vertex as at base, with tubercle at vertex bearing 3 ocelli (Figs. 78I, 80A) 43 Frons parallel-sided or but slightly narrowed below (Figs. 78J, 79A); if strongly narrowed, then callus square or wider than high
43(42). Frons over six times as high as basal width, with strong tubercle at vertex and callus much higher than wide, callus touching eyes (Fig. 78I). Black flies with thorax and abdomen boldly marked with white. (Brazil: Amazonas) <i>Stenotabanus</i> ( <i>Cretotabanus</i> ) Fairchild, 1969
Frons below five times as high as basal width, with slightly raised tubercle at vertex, callus close to but not touching eyes (Fig. 80A). Muscoid-like flies with pale grey and dark-brown scutum and blackish-brown abdomen with a series of large sublateral pale grey spots on tergites 3-6. (Bolivia)
<ul> <li>44(42). Basal frontal callus rarely with upper median prolongation: a median expanded callus never present (Fig. 78J). Frons generally broad, often slightly narrowed below, the basal callus generally as wide as or wider than high (Fig. 78J). Abdomen whitish to brown, patternless, or with median stripe and often with sublateral rows of pale spots. Eye often with 2-3 or more transverse green bands (Southwestern U.S.A. and Neotropical, to southern Brazil)</li></ul>
Basal frontal callus usually with upper median prolongation (Figs. 79A-B), a median expanded callus and black hairs patch often present. Frons generally narrower than above, seldom narrowed below, the basal callus frequently higher than wide, rarely wider than high (Figs. 79A-B). Abdomen variable, but not whitish (Fig. 79F) and rarely with sublateral spots. Eye normally with two green bands (Figs. 79B-C) (U.S.A. (Arizona) and Neotropical, except southern area)
<ul> <li>45(37). Eye bare. Frontal callus replaced by a strong median sulcus (Fig. 80B). Antennal flagellum with only two free flagellomeres (Fig. 80C). (Mexico)</li></ul>
<ul> <li>46(45). Eye bare, bicolored, reddish-violet above, green below. Face with extensive bare areas (Fig. 80D). Wing with apex and large circular spot surrounding all crossveins brown (Fig. 80E). (Southeastern Brazil) Anaerythrops Barretto, 1948 Eye sometimes pilose, unicolorous or with a simple dark stripe. Wing not as above</li></ul>
47(46). Small, black, and almost wholly shining flies, lacking pollinosity even on frons and face (Fig. 81A). Wing entirely hyaline. Female sternite 8 with wide base (Fig. 81E). Male terminalia as in Figs. 82B-C. (Southern Argentina and Chile)
Generally larger flies, variously colored and, even if black, extensively pollinose
48(47). Basal plate of antennal flagellum with a strong, acute, dorsal projection of tooth (Figs. 83B, 84C). Frontal callus ridge-like (Fig. 83A, 84B). Tubercle at vertex well-marked, with vestiges of ocelli (Figs. 83A, 84B). Black flies with all wing avoing beautiful marked, with vestiges of ocelli (Figs. 83A, 84B). Black flies with all

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Chile and Argentina, extending eastwards to Brazil (Rio Grande do Sul) and Uruguay) ... Dasybasis Macquart, 1847

101A.	C. 103B	). Other combinations of characters	57
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Stouter flies, frons not over 6 times as high as wide (Fig. 100I); palpus inflated (Fig. 100J); antenna broader (Fig. 100K); proboscis hardly longer than palpi, labella large. Eye green or brick-red in life. (Panama, Colombia, Peru) ...... *Philipotabanus (Mimotabanus)* Fairchild, 1964b

- 62(61). Wing hyaline or evenly tinted, the costal cell often darker, but never with spots on crossveins or apical clouds. Frontal callus clavate or ridge-like (Figs. 101A, C). Abdomen black or brown, nearly always with transverse bands, at least on tergite 4 (Fig. 101F), rarely otherwise. At least scutellum and often mesonotum pale pollinose and pale haired, generally contrasting with abdomen. Appendix on fork of third vein absent. Eye unbanded, dark. Female: genital furca with base from flat to acute (Fig. 101G); cerci as long as wide or longer than wide (Fig. 101J) and sternite 8 elongate (Fig. 101I). Male gonocoxite rounded apically (Fig. 102E) (Mexico to Panama to northern Argentina)

#### Tribe Tabanini (Figs. 105-109)

#### Key to genera

1. Eye generally pilose in female, always bare in male	2
Eye very rarely sparsely pilose in female, generally bare in male	3

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Figure 1. *Promycteromyia philippii* (Philip, 1958). Male (A-D), female (E-F). A. Abdomen, dorsal view. B. Terminalia, lateral view. C. Epandrium and cerci. D. Aedeagus and gonostyli. E. Spermathecal ducts. F. Tergites 9-10 and cerci. Abbreviations: *aa*, aedeagal apodeme; *ae*, aedeagus; *ba*, basistylus; *ce*, cercus; *di*, dististylus; *Do*, dorsal view; *ea*, endophallic apodeme; *ep*, epandrium; *fa*, flagellar aedeagus; *go*, gonostyli; *hy*, hypoproct; *lf*, lateral flap; *pv*, penis valve; *sc*, sclerotized portion; *st*, style; t 9, tergite 9; *Ve*, ventral view; *vp*, ventral plate of proctiger.



Figure 2. *Promycteromyia derocerca* Coscarón & Philip, 1979. Female (A-H), male (I-M). A. head, lateral view. B. Antenna. C. Palpus. D. head, frontal view. E. External genital appendages, lateral view. E. same, caudal view. G. Genital furca; cd, caudal spermathecal ducts; sc, sclerotized portion (twisted). H. Tergites 9-10 and cerci. I. Terminalia, lateral view. J, same, lateral view. K. Epandrium and cerci. L. Apex of basistylus with style and dististylus. M. Dististylus, ventral view.



Figure 3. *Mycteromyia conica* (Bigot, 1857). Male (A-H), female (J-M). A. Head, frontal view. B, same, lateral view. C. Wing. D. Detail of basicosta. E. Genital appendages, caudal view. F. Same, lateral view. G. Aedeagus and gonostyli. H. Epandrium and cerci. I. Head, lateral view. J. Genital furca and spermathecal ducts. K Sclerotized portion of spermathecal ducts. L. Tergites 9-10 and cerci. M. Sclerite 8 and gonapophyses.



Figure 4A-D. *Silvestriellus flaviventris* (Barretto & Duret, 1954). Male. A. Head, lateral view. B. Same, frontal view. C. Palpus. D. Abdomen, dorsal view. E-G. *Silvestriellus patagonicus* Brèthes, 1910. Female. E. Head, lateral view. F. Same, frontal view. G. Palpus. H-M. *Silvestriellus schlingeri* Coscarón & Philip, 1979. Male (H-J), female (K-M). H. Aedeagus and gonostyli. I. Epandrium and cerci. J. Head, lateral view. K. Genital furca and spermathecal ducts. L. Tergites 9-10 and cerci. M. Sternite 8 and gonapophyses. N-O. *Silvestriellus martinezi* (Barretto & Duret, 1954). Female. N. Head, frontal view. O. Same, lateral view. P. Palpus.



Figure 5. *Apatolestes (Apatolestes) comastes* Williston, 1885. Female. A. Head, frontal view. B. Same, lateral view. C. Frons. D. Palpus. E. Wing.



Figure 6. *Austromyans dasyops* (Philip & Coscarón, 1971). Female. A. Head, lateral view. B. Frons. C. Antenna, D. Palpus. E. Sternite 8 and gonapophyses.



Figure 7. *Protodasyapha (Protodasyapha) hirtuosa* (Philippi, 1865). Female. A. Head, lateral view. B. Same, frontal view. C. Frons. D. Antenna. E. Palpus. F. Genital furca and spermathecal ducts. G. Sternite 8 and gonapophyses. H. Tergites 9-10 and cerci.





Figure 8. *Protodasyapha* (*Protodasyapha*) *hirtuosa* (Philippi, 1865). Male. A. Antenna. B. Palpus. C. Epandrium and cerci. D. Aedeagus and gonostyli. E. Dististylus.


Figure 9. *Protodasyapha* (*Curumyia*) *lugens* (Philippi, 1865). Female. A. Head, frontal view. B. Same, lateral view. C. Frons. D. Palpus. E. Palpal sensory area. F. Macrotrichiae of palpal sensory area. G. Antenna. H. Genital furca and spermathecal ducts. I. Tergites 9-10 and cerci. J. Sternite 8 and gonapophyses.



Figure 10. *Protodasyapha (Curumyia) lugens* (Philippi, 1865). Male. A. Head, frontal view. B. Same, lateral view. C. Antenna. D. Palpus. E. Tip of aedeagus. F. Aedeagus and gonostyli. G. Dististylus, ventral view. H. Same, lateral view. I. Epandrium and cerci.



Figure 11. *Veprius presbiter* Rondani, 1863. Female. A. Head, frontal view. B. Same, lateral view. C. Palpus. D. Antenna. E. Frons. F. Basicosta. G. Genital furca and spermathecal ducts. H. Sternite 8 and gonapophyses. I. Tergites 9-10 and cerci.



Figure 12. *Veprius presbiter* Rondani, 1863. Male. A. Head, frontal view. B. Same, lateral view. C. Antenna. D. Palpus. E. Aedeagus and gonostyli. F. Dististylus. G. Tip of aedeagus. H. Epandrium and cerci.



Figure 13. *Zophina eiseni* (Townsend, 1895). Female (A-E), male (F-J). A. Frons. B. Head, lateral view. C. Antenna. D. Palpus. E. Genital furca and spermathecal ducts. F. Head, lateral view. G. Same, frontal view. H. Antenna. I. Aedeagus and gonostyli. J. Epandrium and cerci.

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Figure 14. *Fairchildimyia penai* Philip & Coscarón, 1971. Female. A. Head, lateral view. B. Frons. C. Palpus. D. Sternite 8 and gonapophyses.



Figure 15 A-J. *Fairchildimyia penai* Philip & Coscarón, 1971. Male. A. Head, frontal view. B. Same, lateral view. C. Wing. D. Abdomen, dorsal view. E. Palpus. F. Antenna. G. Epandrium and cerci. H. Aedeagus and gonostyli. I. Dististylus. J-L. *Fairchildimyia mendozana* (Enderlein, 1925). Female. J. Antenna. K. Palpus. L. Palpal sensory area.

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Figure 16 *Chaetopalpus annulicornis* (Philippi, 1865). Female. A. Head, frontal view. B. Same, lateral view. C. Antenna. D. Palpus. E. Frons. F. Abdomen, dorsal view. G. Basicosta. H. Palpal sensory area. I. Same (higher magnification). J. Genital furca and spermathecal ducts. K. Apex of spermatheca. L. Tergites 9-10 and cerci. M. Sternite 8 and gonapophyses.



Figure 17. *Chaetopalpus annulicornis*(Philippi, 1865). Male. A. Head, frontal view. B. Same, lateral view. C. Antenna. D. Palpus. E. Aedeagus and gonostyli. F. Dististylus, ventral view. G. Same, dorsal view. H. Epandrium and cerci.



Figure 18. *Archeomyotes angustipennis* Philip & Coscarón, 1971. Female. A. Head, lateral view. B. Frons. C. Antenna. D. Palpus. E. Wing. F. Genital furca and spermathecal ducts.



Figure 19. A. *Esenbeckia (Proboscoides) dichroa* (Brèthes, 1910). Female. Head, frontal view. B-D. *Esenbeckia (Palassomyia) fascipennis* (Macquart, 1838). Female. B. Head, frontal view. C. Same, lateral view. D. Wing. E-F. *Esenbeckia (Ricardoa) subguttata* Fairchild, 1964. Female. E. Head, lateral view. F. Palpus.



Figure 20. *Esenbeckia (Esenbeckia) filipalpis* (Williston, 1895). Female (A-I, male (J-L). A. Head, frontal view. B. Head, lateral view. C. Antenna. D-E. Palpus. F. Abdomen. G. Sternite 8 and gonapophyses. H. Tergites 9-10 and cerci. I. Genital furca and spermathecae. J. Aedeagus and gonostyli. K. Dististylus, ventral view. L. Epandrium and cerci.



Figure 21. A-B. *Protosilvius sp.* A. Head, lateral view. B. Antenna. C-F. *Protosilvius phoeniculus* Fairchild, 1962. Male (C-E), female (F-G). C. Antenna. D. Male terminalia, ventral view. E. Tergite 9 and cerci. F. Frons. G. Sternite 8.



Figure 22. *Scepis nivalis* Walker, 1850. Female. A. Head, frontal view. B. Same, lateral view. C. Palpus. D. Antenna. E. Sternite 8 and gonapophyses. F. Tergite 10 and cerci. G. Genital furca and spermathecae. H. Detail of proximal section of spermathecal duct.



Figure 23. *Scepsis nivalis* Walker, 1850. Male. A. Head, anterior view. B. Same, lateral view. C. Antenna. D. Palpus. E. Epandrium and cerci. F. Aedeagus and gonostyli. G. Dististylus, dorsal view.



Figure 24. A-E. *Caenopangonia hirtipalpis* (Bigot, 1892). Male. A. Head, frontal view. B. External genital appendages, Caudal view. C. Epandrium and cerci. D. Aedeagus and gonostyli. E. Dististylus, ventral view. F-J. *Caenopangonia brevirostris* (Philippi, 1865). Female. F. Head, frontal view. G. Palpus. H. Sternite 8 and gonapophyses. I. Genital furca and spermathecal ducts. J. Tergites 9-10 and cerci. K-N. *Caenopangonia aspera* (Philip, 1958). Male. K. Head, frontal view. L. Palpus. M. Head, lateral view. N. Aedeagus and gonostyli.



Figure 25. A-E. *Scaptia* (*Scaptia*) *collaris* (Philippi, 1865). Female (A-B, D-E), male (C). A. Head, frontal view. B. Frons. C. Palpus. D. Antenna. E. Palpus. F-O. *Scaptia* (*Scaptia*) *lata* (Guérin-Méneville, 1835). Female (F-L), male (M-O). F. Palpus. G. Tergites 9-10 and cerci. H. Spermatheca. I. Frons. J. Antenna. K. Sternite 8 and gonapophyses. L. Genital furca and spermathecal ducts. M. Epandrium and cerci. N. Dististylus. O. Aedeagus and gonostyli.



Figure 26. *Scaptia* (*Lepmia*) *seminigra* (Ricardo, 1902). Female. A. Head, lateral view. B. Frons. C. Palpus. D. Tergites 9-10 and cerci. E. Sternite 8 and gonapophyses. F. Genital furca and spermathecae.



Figure 27. *Scaptia (Lepmia) seminigra* (Ricardo, 1902). Male. A. Head, lateral view. B. Palpus. C. Gonocoxite, basistyli, dististyli and aedeagus. D. Dististylus. E. Epandrium, ventral view of proctiger and cerci.



Figure 28. *Scaptia (Pseudomelpia) horrens* Enderlein, 1925. A. Head, lateral view. B. Frons. C. Antenna. D. Palpus. E. Abdomen, dorsal view.



Figure 29. *Scaptia (Pseudoscione) stictica* Wilkerson & Coscarón, 1984. A. Frons. B. Head of female, lateral view. C. Palpus of female. D. Tergites 9-10 of female. E. Head of male, lateral view. F. Genital furca and spermathecal ducts. G. Infra-anal plate. H. Male epandrium and cerci. I. Male gonopod and aedeagus.



Figure 30. *Scione flavohirta* Ricardo, 1902. Female. A. Antenna. B. Abdomen, dorsal view. C. Frons. D. Head, lateral view. E-F. Palpus. G. Wing. H. Sternite 8 and gonapophyses. I. Hypoproct, tergites 9-10 and cerci. J. Genital furca and spermathecal ducts.



Figure 31. *Scione flavohirta* Ricardo, 1902. Male. A. Palpus. B. Aedeagus, basistyli and dististyli. C. Dististylus. D. Epandrium, hypandrium and cerci.

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Figure 32. *Fidena (Neopangonia) pusilla* (Lutz, 1909). Female. A. Head, lateral view. B. Frons. C. Palpus. D. Mesonotum, dorsal view. E. Wing. F. Genital furca and spermathecal ducts. .G. Tergites 9-10 and cerci. H. Sternite 8 and gonapophyses.



Figure 33. *Fidena (Laphriomyia) mirabilis* (Lutz, 1911). Female. A. Head, lateral view. B. Frons. C. Palpus. D. Hind leg. E. Genital furca and spermathecal ducts. F. Tergites 9-10 and cerci. G. Sternite 8 and gonapophyses.



Figure 34. *Fidena* (*Leptofidena*) *morio* (Wulp, 1881). A-F. Female. A. Head, lateral view. B. Frons. C. Palpus. D. Genital furca and spermathecal ducts. E. Tergites 9-10 and cerci. F. Sternite 8 and gonapophyses. G-I. Male. G. Head, lateral view. H. Palpus. I. Dististylus.



Figure 35. *Fidena (Fidena) erythronotata* (Bigot, 1892). Female. A. Head, lateral view. B. Frons. C. Palpus. D. Tergites 9-10 and cerci. E. Sternite 8 and gonapophyses. F. Genital furca and spermathecal ducts.



Figure 36. *Fidena (Fidena) neglecta* Kröber, 1931. Male. A. Head, lateral view. B. Palpus. C. Basistyli, dististyli and aedeagus. D. Dististylus. E. Epandrium, cerci and proctiger.



Figure 37. A-B. *Pityocera (Pityocera) festai* Giglio-Tos, 1896. A. Antenna. B. Palpus. C-D. *Pityocera (Pseudelaphella) nana* (Walker, 1850). C. Antenna. D. Genital-furca and spermarhecal ducts.



Figure 38. A-D. Pityocera (Elaphella) cervus (Wiedemann, 1828). A. Head, lateral view. B. Palpus. C. Frons. D. Antenna.



Figure 39. A-B. *Pseudotabanus (Coracella ) carbo* (Macquart, 1850). Female. A. Frons. B. Antenna. C-G. *Pseudotabanus (Coracella) araucana* (Coscarón, 1972). Female (C-F). C. Head, frontal view. D. Genital furca and spermathecal ducts. E. Sternite 8 and gonapophyses. F. Tergites 9-10 and cerci. G. Male aedeagus and gonostyli.



Figure 40. *Betrequia ocellata* Oldroyd, 1970. Female. A. Head, lateral view. B. Wing. C. Genital furca and spermathecal ducts. D. Sternite 8 and gonapophyses. E. Tergites 9-10 and cerci.



Figure 41. A-D. *Chrysops crucians* Wiedemann, 1828. Female. A. Head, lateral view. B. Frons. C. Wing. D. Abdomen, dorsal view. E-I. *Chrysops flavoscutellatus* Kröber, 1926. Female. E. Head, lateral view. F. Frons. G Wing. H. Abdomen. I. Genital furca and spermathecal ducts. J-O. *Chrysops flinti* Coscarón, 1979. Female. J. Head, lateral view. K. Frons. L. Antenna. M. Wing. N. Abdomen, dorsal view. O. Genital furca and spermahecal ducts.



Figure 42. *Chrysops laetus* Fabricius, 1805. A. Basistyli, aedeagus and dististyli. B. Dististylus. C. Paraprocts, cerci and hypoproct.



Figure 43. *Silvius (Griseosilvius quadrivittatus* (Say, 1823). Female. A. Head, frontal view. B. Antenna. C. Wing. D. Abdomen. E. Genital furca and spermathecal ducts. F. Sternite 8 and gonapophyses. G. Tergites 9-10 and cerci.



Figure 44. Silvius (Assipala) melanopterus (Hine, 1905). A. Head, frontal view. B. Same, lateral view.


Figure 45. *Bolbodimyia lateralis* Kröber, 1930. Female. A. Head, frontal view. B. Same, lateral view. C. Frons. D. Antenna. E. Palpus. F. Wing.

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Figure 46. *Bolbodimyia lateralis* Kröber, 1930. Male. (A-B), female (C-F). A. Aedeagus and gonostyli. B. Dististylus. C. Genital furca and spermathecal ducts. D. Spermatheca. F. Sternite 8 and gonapophyses. F. Tergites 9-10 and cerci.



Figure 47. A-B. *Acanthocera* (*Querbetia*) *chaineyi* Fairchild & Burger, 1994. Female. A. Head, lateral view. B. Same, frontal view. C. *Acanthocera* (*Querbetia*) *inopinata* (Fairchild, 1972). Female. Antenna. D. *Holcopsis fenestrata* Enderlein, 1925. Female. Frons.



Figure 48. *Chlorotabanus parviceps* (Kröber, 1934). Female. A. Head, lateral view. B. Frons. C. Antenna. D. Palpus. E. Genital furca and spermathecal ducts. F. Spermatheca. G. Sternite 8 and gonapophyses. H. Tergites 9-10 and cerci.



Figure 49. *Chlorotabanus parviceps* (Kröber, 1934). Male. A. Head, lateral view. B. Palpus. C. Aedeagus and gonostyli. D. Dististylus. E. Epandrium and cerci.



Figure 50. *Pachyschelomyia notopleuralis* Barretto, 1950. Female. A. Head, frontal view. B. Same, lateral view. C. Frons. D. Antenna. E. Palpus. F. Wing. G. Genital furca and spermathecal ducts. H. Spermatheca. I. Sternite 8 with gonapophyses. J. Tergites 9-10 and cerci.



Figure 51. *Myiotabanus barrettoi* Fairchild, 1971. Male (A-F), female (G-N). A. Frons. B. Antenna. C. Palpus. D. Sternite 8 and gonapophyses. F. Genital furca and spermathecal ducts. F. Tergites 9-10 and cerci. G. Head, frontal view. H. Same, lateral view. I. Antenna. J. Palpus. K. Thorax and abdomen, dorsal view. L. Aedeagus and gonostyli. M. Dististylus. N. Epandrium and cerci.



Figure 52. A-K. *Phaeotabanus limpidapex* (Wiedemann, 1828). Female (A-I), male (J-K). A. Head, frontal view. B. Frons. C. Antenna. D. Palpus. E. Wing. F. Sternite 8 and gonapophyses. G. Genital furca and spermathecal ducts. H. Spermatheca. I. Tergites 9-10 and cerci. J. Head, lateral view. K. Palpus. L-O. *Eristalotabanus violaceus* Kröber, 1931. Female. L. Head, lateral view. M. Palpus. N. Frons. O. Antenna.

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Figure 53. Acanthocera (Mimodynerus) anacantha Lutz & Neiva, 1915. Female. A. Head, frontal view. B. Same, dorsal view. C. Frons. D. Palpus. E. Thorax and abdomen, dorsal view. F. Abdomen, dorsal view.



Figure 54. A-C. Acanthocera (Acanthocera) coarctata (Wiedemann, 1828). Female. A. Head, lateral view. B. Same, frontal view. C. Wing. D. D-F. Acanthocera (Acanthocera) marginalis Walker, 1854. Female. D. Head, frontal view. E. Same, dorsal view. F. Palpus. G. Acanthocera (Polistimima) polistiformis Fairchild, 1961. Female. Antenna.



Figure 55. A-H. *Acanthocera* (*Acanthocera*) exstincta (Wiedemann, 1828). Female. A. Frons. B. Head, lateral view. C. Antenna. D. Palpus. E. Head, thorax and abdomen, dorsal view. F. Wing. G. Genital furca and spermathecal ducts. H. Sternite 8 and gonapophyses. I-O. *Acanthocera* (*Acanthocera*) coarctata (Wiedemann, 1828). Female. I. Frons. J. Antenna. K. Palpus. L. Head, thorax and abdomen, dorsal view. M. Tergites 9-10, cerci and hypoproct. N. Sternite 8 and gonapophyses. O. Distal portion of spermatheca.

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Figure 56. *Dichelacera* (*Dichelacera*) *fuscipes* Lutz, 1915. Female. A. Head, frontal view. B. Frons. C. Antenna. D. Palpus. E. Wing. F. Genital furca and spermathecal ducts. G. Spermatheca. H. Sternite 8 and gonapophyses. I. Tergites 9-10 and cerci.



Figure 57. *Dichelacera* (*Dichelacera*) *fuscipes* Lutz, 1915. Male. A. Head, frontal view. B. Same, lateral view. C. Palpus. D. Aedeagus and gonostyli. E. Dististylus. F. Epandrium and cerci.



Figure 58. A-C. Dichelacera (Desmatochelacera) transposita Walker, 1854. Female. A. Frons. B. Antenna. C. Wing. D-F. Dichelacera (Idiochelacera) subcallosa Fairchild & Philip, 1960. Female. D. Frons. E. Palpus. F. Antenna. G-I. Dichelacera (Dichelacera) boliviensis (Brèthes, 1910). Female. G. Frons. H. Head, lateral view. I. Antenna. J. Acanthocera (Nothocanthocera) tenuicornis (Lutz, 1915). Female. Wing. K-N. Dichelacera (Orthostyloceras) ambigua (Lutz & Neiva, 1914). Female. K. Frons. L. Antenna. M. Palpus. N. Wing.



Figure 59. A-E. *Catachlorops (Psalidia) conspicuus* (Lutz & Neiva, 1914). Female. A. Head, frontal view. B. Same, dorsal view. C. Proboscis, lateral view. D. Antenna. E. Palpus. F. Wing. G-J. *Catachlorops (Psalidia) furcatus* (Wiedemann, 1828. Female. G. Frons. H. Antenna. I. Palpus. J. Wing.



Figure 60. *Catachlorops (Catachlorops) leptogaster* Barretto, 1946. Female. A. Head, frontal view. B. Same, lateral view. C. Frons. D. Antenna. E. Palpus. F. Wing. G. Sternite 8 and gonapophyses. H. Genital furca and spermathecal ducts. I. Spermatheca. J. Tergites 9-10 and cerci.



Figure 61. Catachlorops (Rhamhidommia) muscosus (Enderlein, 1925). Female. A. Frons. B. Antenna. C. Wing.



Figure 62. A-B. *Catachlorops (Psarochlorops) difficilis* (Kröber, 1931). Female. A. Frons. B. Wing. C-D. *Catachlorops (Psarochlorops) testaceus (Macquart, 1846). Female. C. Head, dorsal view. D. Same, frontal view. E-H. Catahlorops (Hadrochlorops) unicolor (Lutz, 1912). Female. E. Head, frontal view. F. Proboscis. G. Palpus. H. Antenna.* 



Figure 63. *Catachlorops (Amphichlorops) flavus (Wiedemann, 1828).* Female. A. Head, frontal view. B. Proboscis. C. Antenna. D. Palpus.



Figure 64. A-G. *Stibasoma* (*Stibasoma*) *theotaenia* (Wiedemann, 1828). Female. A. Head, lateral view. B. Frons. C. Antenna. D. Palpus. E. Genital furca and spermathecal ducts. F. Sternite 8 and gonapophyses. G. Tergites 9-10 and cerci.



Figure 65. *Stibasoma (Stibasoma) theotaenia* (Wiedemann, 1828). Male. A. Palpus. B. Wing. C. Hind leg. D. Basistyli, dististyli and aedeagus. E. Cerci and paraprocts. [Figs. D-E with the same scale].



Figure 66. *Cryptotylus unicolor* (Wiedemann, 1828). Female. A. Head, lateral view. B. Frons. C. Antenna. D. Palpus. E. Genital furca and spermathecal ducts. F. Spermatheca. G. Tergites 9-10 and cerci. H. Sternite 8 and gonapophyses.



Figure 67. *Cryptotylus unicolor* (Wiedemann, 1828). Male. A. Head, lateral view. B. Same, frontal view. C. Antenna. D. Palpus. E. Aedeagus and gonostyli. F. Dististylus. G. Epandrium and cerci.



Figure 68. A. *Dasychela* (*Triceratomyia*) *macintyrei* (Bequaert, 1937). Female. Antenna. B-E. *Dasychela* (*Dasychela*) *sp.* Female. C. Genital furca. D. Sternite 8 and gonapophyses. E. Tergites 9-10 and cerci.



Figure 69. *Lepiselaga (Lepiselaga) crassipes* (Fabricius, 1805). Female. A. Frons. B. Antenna. C. Palpus. D. Wing. E. Hind leg. F. Genital furca and spermathecal ducts. G. Spermatheca. H. Sternite 8 and gonapophyses. I. Tergites 9-10 and cerci.

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Figure 70. *Lepiselaga (Lepiselaga) crassipes (Fabricius, 1805)*. Male. A. Palpus. B. Epandrium and cerci. C. Aedeagus and gonostyli.



Figure 71. *Lepiselaga (Conoposelaga) albitarsis* Macquart, 1850. Female. A. Head, lateral view. B. Same, frontal view. C. Antenna. D. Palpus. E. Wing. F. Genital furca and spermathecal ducts. G. Spermatheca. H. Sternite 8 and gonapophyses. I. Tergite 10, cerci and hypoproct.



Figure 72. Lepiselaga (Conoposelaga) albitarsis Macquart, 1850. Male. A. Palpus. B. Aedeagus and gonostyli.



Figure 73. A. *Oopelma globicornis* (Wiedemann, 1821). Antenna. B-D. *Himantostylus intermedius* Lutz, 1913. Female. B. Antenna. C. Palpus. D. Sternite 8 and gonapophyses.



Figure 74. *Erioneura fuscipennis* (Wiedemann, 1828). Female. A. Frons. B. Antenna. C. Palpus. D. Wing. E. Genital furca and spermatheca. F. Sternite 8 and gonapophyses. G. Tergites 9-10 and cerci.



Figure 75. *Stigmatophthalmus altivagus* Lutz, 1913. Female (A-E), male (F-G). A. Antenna. B. Wing. C. Sternite 8 and gonapophyses. D. Tergites 9-10 and cerci. E. Genital furca and spermathecae. F. Aedegus and gonostyli. G. Epandrium and cerci.



Figure 76. *Eutabanus pictus* Kröber, 1930. Female. A. Head and thorax, dorsal view. B. Frons. C. Wing. D. Genital furca and spermathecae. E. Sternite 8 and gonapophyses.



Figure 77. *Diachlorus flavitaenia* Lutz, 1913. Female. A. Head, lateral view. B. Frons. C. Antenna. D. Palpus. E. Thorax and abdomen, dorsal view. F. Genital furca and spermathecal ducts. G. Spermatheca. H. Sternite 8 and gonapophyses. I. Tergites 9-10 and cerci.



Figure 78. *Stenotabanus (Brachytabanus) platyfrons* Fairchild, 1964. Female. A. Head, frontal view. B. Same, lateral view. C. Antenna. D. Abdomen, dorsal view. E. Sternite 8 and gonapophyses. F. Genital furca and spermathecal ducts. G. Spermatheca. H. Tergites 9-10 and cerci. I. *Stenotabanus (Cretotabanus) cretatus* Fairchild, 1961. Female. Frons. J-K. *Stenotabanus (Aegialomyia) littoreus* (Hine, 1907). Female. J. Frons. K. Antenna. L. *Stenotabanus (Wilkersonia) roxannae* Wikerson, 1979. Female. Frons.



Figure 79. A-I. *Stenotabanus (Stenotabanus) incipiens* (Walker, ). A-J. Female. A. Frons. B. Head, frontal view. C. Head, lateral view. D. Antenna. E. Palpus. F. Abdomen, dorsal view. G. Sternite 8 and gonapophyses. H. Genital furca. I. Apical portion of spermatheca. J. Cerci, tergites 9-10 and hypoprot. K-S. Male. K. Head, frontal view. L. Head, lateral view. M. Antenna. N. Palpus. O. Abdomen, dorsal view. P. Basistyli, dististyli and aedeagus. Q. Dististylus. R. Apical portion of aedeagus. S. Paraprocts and hypoprot.



Figure 80. A. *Roquezia signifera* Wilkerson, 1985. Female. Frons. B-C. *Teskeyellus hirsuticornis* Philip & Fairchild, 1974. Female. B. Frons. C. Antenna. D-E. *Anaerythrops lanei* Barretto, 1948. Female. D. Head, frontal view. E. Wing. F. *Hemichrysops fascipennis* Kröber, 1930. Female. Frons. G. *Spilotabanus multiguttatus* (Kröber, 1930). Frons.


Figure 81. *Scaptiodes gagatina* (Philippi, 1865). Female. A. Frons. B. Antenna. C. Palpus. D. Genital furca and gonapophyses. E. Sternite 8 and gonapophyses. F. Tergites 9-10 and cerci. G. Hypoproct. H. Spermatheca.

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Figure 82. Scaptiodes gagatina (Philippi, 1865). Male. A. Palpus. B. Aedeagus and gonostyli. C. Epandrium and cerci.



Figure 83. *Nubiloides nigripennis* (Philippi, 1865). Female. A. Frons. B. Antenna. C. Palpus. D. Genital furca and spermathecal ducts. E. Sternite 8 and gonapophyses. F. Tergites 9-10 and cerci. G. Spermatheca. H. Hypoproct.



Figure 84. *Nubiloides schajovskoi* (Coscarón, 1974). Female. A. Head, lateral view. B. Frons. C. Antenna. D. Palpus. E. Abdomen, dorsal view. F. Genital furca and spermathecal ducts. G. Sternite 8 and gonapophyses. H. Tergites 9-10 and cerci.



Figure 85. *Haematopotina argentina* (Brèthes, 1910). Female. A. Frons. B. Head, lateral view. C. Antenna. D. Palpus. E. Wing. F. Head, thorax and abdomen, dorsal view. G. Genital furca and spermathecal ducts. H. Spines of upper portion of furca. I. Spermatheca. J. Hypoproct. K. Sternite 8 and gonapophyses. L. Tergites 9-10 and cerci.



Figure 86. *Haematopotina argentina* (Brèthes, 1910). Male. A. Aedeagus. B. Palpus. C. Aedeagus and gonostyli. D. Epandrium and cerci.



Figure 87. *Agelanius lanei* (Coscarón & Philip, 1967). Female. A. Frons. B. Antenna. C. Palpus. D. Head, thorax and abdomen, dorsal view. E. Genital furca and spermathecal ducts. F. Hypoproct. G. Sternite 8 and gonapophyses. H. Tergites 9-10 and cerci.



Figure 88 Agelanius lanei (Coscarón & Philip, 1967). Male. A. Antenna. B. Palpus. C. Aedeagus and gonostyli. D. Epandrium and cerci.



Figure 89. *Acellomyia fontanensis* (Coscarón, 1962). Female. A. Thorax and abdomen, dorsal view. B. Palpus. C. Head, dorsal view. D. Head, lateral view. E. Antenna.



Figure 90. *Acellomyia fontanensis* (Coscarón, 1962). Female. A. Wing. B. Genital furca and spermathecal ducts. C. Comb on distal portion of genital furca. D. Tip of spermatheca. E. Caudal end of spermathecal duct. F. Hypoproct. G. Tergite 9, basal plate of tergite 10 and cerci. H. Sternite 8 and gonapophyses.



Figure 91 *Acellomyia albohirta* (Walker, 1837). Male. A. Head, frontal view. B. Antenna. C. Aedeagus and gonostyli. D. Epandrium and cerci.



Figure 92. *Acellomyia albohirta* (Walker, 1837). Female. A. Head, thorax and abdomen, dorsal view. B. Frons. C. Antenna. D. Palpus. E. Sternite 8 and gonapophyses. F. Genital furca and spermathecal ducts. G. Tergites 9-10 and cerci. H. Hypoproct. I. Spermatheca.



Figure 93. *Dasybasis albosignata* (Kröber, 1930). Female. A. Frons. B. Head, lateral view. C. Head, thorax and abdomen, dorsal view. D. Antenna. E. Palpus. F. Genital furca and spermathecal ducts. G. Hypoproct. H. Sternite 8 and gonapophyses. I. Tergites 9-10 and cerci.



Figure 94. *Dasybasis testaceomaculata* (Macquart, 1838). Female. A. Frons. B. Antenna. C. Palpus. D. Spermatheca. E. Detail of spermathecal wall. F. Sternite 8 and gonapophyses. G. Tergites 9-10 and cerci.



Figure 95. *Dasybasis (Dasybasis) testaceomaculata* (Macquart, 1838). Male. A, C. Antenna, showing variation. B, D. Palpus, variation. E. Aedeagus and gonostyli. F. Epandrium and cerci.



Figure 96. *Selasoma tibiale* (Fabricius, 1805). Female. A. Frons. B. Antenna. C. Palpus. D. Wing. E. Hind leg. F. Genital furca and spermathecal duct. G. Tergites 9-10 and cerci. H. Sternite 8 and gonapophyses.



Figure 97. *Pseudacanthocera brevicornis* (Enderlein, 1925). Female. A. Head, lateral view. B. Frons. C. Antenna. D. Palpus. E. Thorax and abdomen, dorsal view. F. Genital furca and spermathecal ducts. G. Spermatheca. H. Sternite 8 and gonapophyses. I. Tergites 9-10 and cerci.

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Figure 98. *Dicladocera (Dicladocera) bellicosa.* (Brèthes, 1910). Female. A. Frons. B. Antenna. C. Palpus. D. Wing. E. Basicosta. F. Genital furca and spermathecal ducts. G. Spermathecae. H. Tergites 9-10 and cerci. I Sternite 8 and gonapophyses.

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Figure 99. *Dicladocera (Dicladocera) nubipennis* (Rondani, 1864). Male. A. Frons. B. Antenna. C. Palpus. D. Cerci, paraprocts and hypoproct. E. Basistyli, dististyli and aedeagus.



Figure 100. A-D. Leptapha fumata (Wiedemann, 1821). Female. A. Frons. B. Antenna. C. Palpus. D. Sternite 8 and gonapophyses. E-G. Philipotabanus (Philipotabanus) magnificus (Kröber, 1934). Female. E. Frons. F. Palpus. G. Wing. H. Philipotabanus (Melasmatabanus) fascipennis (Macquart, 1846). Female. Wing. I-K. Philipotabanus (Mimotabanus) fucosus Fairchild, 1958. Female. I. Frons. J. Palpus. K. Antenna.



Figure 101. *Leucotabanus procallosus* Lutz, 1912. Female. A. Head, frontal view. B. Same, lateral view. C. Frons. D. Antenna. E. Palpus. F. Abdomen, dorsal view. G. Genital furca and spermathecal ducts. H. Spermatheca. I. Sternite 8 and gonapophyses. J. Tergites 9-10 and cerci.



Figure 102. *Leucotabanus procallosus* Lutz, 1912. Male. A. Head, frontal view. B. Same, lateral view. C. Antenna. D. Palpus. E. Aedeagus and gonostyli. F. Dististylus. G. Epandrium and cerci.



Figure 103. *Stypommisa rubrithorax* (Macquart, 1838). Female. A. Head, lateral view. B. Frons. C. Antenna. D. Palpus. E. Abdomen, dorsal view. F. Genital furca and spermathecal ducts. G. Spermatheca. H. Sternite 8 and gonapophyses. I. Tergites 9-10 and cerci.

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Figure 104. *Stypommisa rubrithorax* (Macquart, 1838). Male. A. Head, lateral view. B. Palpus. C. Aedeagus and gonostyli. D. Dististylus. E. Epandrium and cerci.



Figure 105. *Atylotus vargasi* Philip, 1954. Female. A. Frons. B. Genital furca and spermathecae. C. Sternite 8 and gonapophyses. D. Tergites 9-10 and cerci.



Figure 106. *Poeciloderas lindneri* (Kröber, 1930). Female (A-E), male (F-G). A. Head, lateral view. B. Antenna. C. Frons. D. Genital furca and spermathecal ducts. E. Sternite 8 and gonapophyses. F. Aedeagus and gonostyli. G. Epandrium and cerci.



Figure 107. *Phorcotabanus cinereus* (Wiedemann, 1821). Female (A-H), male (I). A. Head, frontal view. B. Same, lateral view. C. Frons. D. Antenna. E. Abdomen, dorsal view. F. Palpus. G. Sternite 8 and gonapophyses. H. Genital furca and spermathecal ducts. I. Aedeagus and gonostyli.



Figure 108. *Tabanus sorbillans* Wiedemann, 1828. Female. A. Head, lateral view. B. Frons. C. Antenna. D. Palpus. E. Abdomen, dorsal view. F. Sternite 8 and gonapophyses.



Figure 109. *Tabanus triangulum* Wiedemann, 1828. Female (A-B, D-G), male (C, H). A. Frons. B-C. Antenna. D. Palpus. E. Abdomen, dorsal view. F. Genital furca and spermathecal ducts. G. Tergites 9-10 and cerci. H. Aedeagus and gonostyli.